

Methods in Litter Decomposition

Time Table

	Morning	Afternoon
Monday 13 rd April	<p>(9:00 – 10:00) Welcome; Lecture 1. Litter decomposition in streams (MAG) LR <i>Coffee break</i></p> <p>(10:30 – 11:15) Lecture 2. Litter quality and environmental quality influences on litter decomposition (MAG) LR</p> <p>(11:15 – 12:45) Lecture 3. Litter decomposition experiments: mass loss determination (VF) LR</p>	<p>(14:30 – 15:15) Lecture 4. Aquatic hyphomycetes: diversity and functions (EC) LR <i>Break</i></p> <p>A → (15:45 – 17:15) Lab. Isolation of aquatic hyphomycetes Part 1 (EC) MicLab</p> <p>B → (15:45 – 17:15) Lab. Phenols, toughness and SLA determination (MAG) ChLab.I <i>Break</i></p> <p>A → (17:45 – 19:15) Lab. Phenols, toughness and SLA determination (MAG) ChLab.I</p> <p>B → (17:45 – 19:15) Lab. Isolation of aquatic hyphomycetes Part 1 (EC) MicLab</p>
Tuesday 14 th April	<p>(9:00 – 9:45) Lecture 5. Fungal biomass (MOG) LR <i>Coffee break</i></p> <p>A → (10:15 – 13:00) Lab. Fungal sporulation: filtration, spores counting and identification (EC) 15°C Lab, ChLab.I & MicLab</p> <p>B → (10:15 – 13:00) Lab. Fungal biomass: ergosterol extraction (MOG) ChLab.I</p>	<p>A → (14:00 – 17:00) Lab. Fungal biomass: ergosterol extraction (MOG) ChLab.I</p> <p>B → (14:00 – 17:00) Lab. Fungal sporulation: filtration, spores counting and identification (EC) 15°C Lab, ChLab.I & ID</p>
Wednesday 15 th April	<p>(9:00 – 9:45) Lecture 6. Fungal sporulation: calculations (EC) LR <i>Coffee break</i></p> <p>A → (10:15 – 12:00) Lab. Isolation of aquatic hyphomycetes Part 2 (EC) MicLab</p> <p>B → (10:15 – 12:00) Lab. Phosphorus determination from litter (VF) ChLab.I</p>	<p>(14:00 – 15:00) Lecture 7. Fungal biomass: HPLC, calculation and discussion (MOG) LR <i>Break</i></p> <p>A → (15:30 – 17:30) Lab. Phosphorus determination from litter (VF) ChLab.I</p> <p>B → (15:30 – 17:30) Lab. Isolation of aquatic hyphomycetes Part 2 (EC) MicLab</p>
Thursday 16 th April	<p>(9:00 – 9:45) Lecture 8. Fungal diversity in freshwaters (CP) LR <i>Coffee break</i></p> <p>(10:15 – 11:00) Lecture 9. Molecular techniques for studying the ecology of aquatic fungi (FC) LR</p> <p>A and B → (11:30-13:00) Lab. DNA extraction (FC, CP, SD)</p>	<p>A and B → (14:00-18:00) Lab. PCR; run agarose gel; DGGE gel preparation and sample loading (CP, FC, SD)</p>
Friday 17 th April	<p>A and B → (9:00 – 10:00) Lab. Gel staining and image acquisition (CP, FC, SD) <i>Coffee break</i></p> <p>A and B → (10:30 – 12:30) TP. Software tools for gel analysis; clusters and ordination analyses of fungal communities (CP, FC, SD) LR</p>	<p>A and B → (13:30 – 15:30) Lab. DNA extraction and amplification (CP, FC, SD) Microbiology</p> <p>A + B (16:00 – 17:00) Lab. Data analysis and interpretation (CP, FC, SD) LR</p> <p>(17:15 – 18:00) Lecture 10. Conclusion: global data interpretation (MAG) LR</p>

CP – Cláudia Pascoal; **EC** – Eric Chauvet; **FC** – Fernanda Cássio; **MAG** – Manuel Graça; **MOG** – Mark Gessner; **SD** – Sofia Duarte; **VF** – Verónica Ferreira. **ChLab.I**– Chemistry Laboratory I; **LR** – Lecture room (Chemistry Department building, 2nd. Floor); **15°C Lab**- Laboratory at 15°C.; **ChLab.II**– Chemistry Laboratory II; **Microbiology** – Laboratory of microbiology; **MicLab** – Microscopy Laboratory. A → & B → two working groups